



KATHOLIEKE UNIVERSITEIT  
**LEUVEN**

 **JÜLICH**  
FORSCHUNGSZENTRUM



# CHARACTERIZATION OF THERMOREVERSIBLE GELS COMPOSED OF MONODISPERSE RODLIKE PARTICLES: RHEOLOGY AND SCATTERING

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# Introduction

*Gels, under flow are liquid,  
but behave like solid at rest*

## Properties

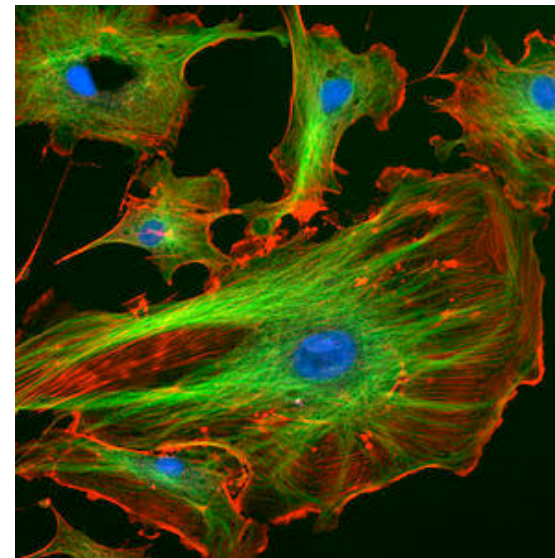
- high moduli
- yield stress
- long relaxation times
- shear thinning
- thixotropy



Food



Paint



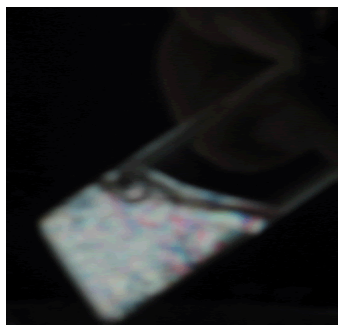
Eukaryotic cytoskeleton

<http://en.wikipedia.org/wiki/>

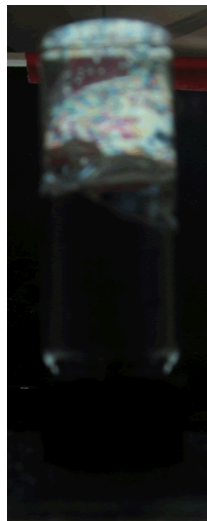
# Introduction

## Advantages of gels composed of rod-like particles

- Relatively low volume fraction needed to form a gel
- Different phases (isotropic, nematic, smectic, columnar)
- Higher gel strength (compared to that made of spheres)
- Relevant to biological cells (actin, protein filaments)

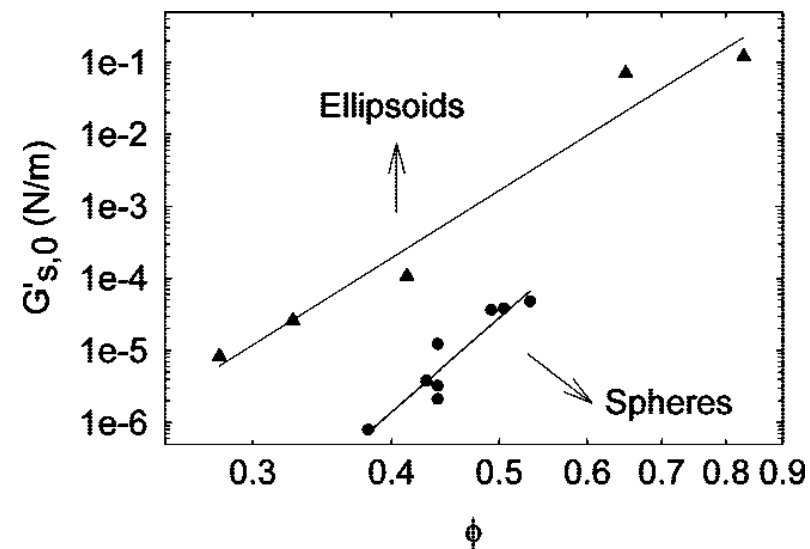


Nematic  
fluid



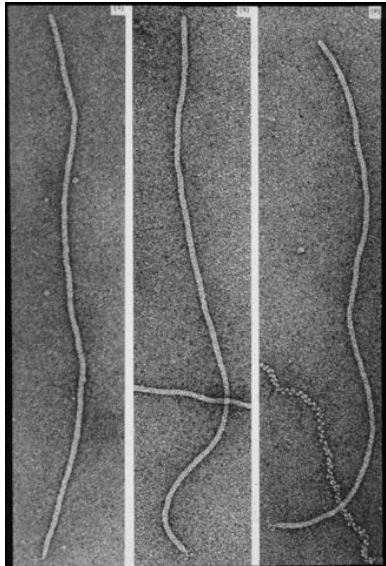
Nematic  
gel

Zhang *et al.* Langmuir, 2009, 25 (4), pp 2437

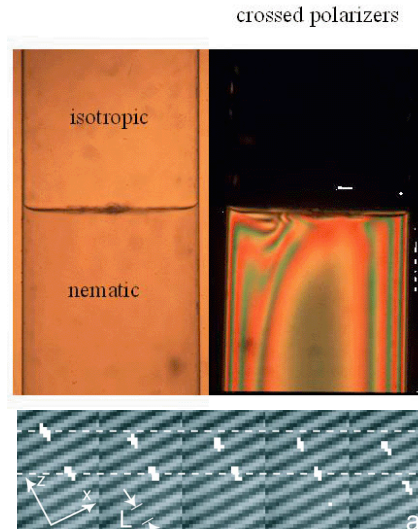


Basavaraj *et al.* Langmuir, 2009, 25 (5), pp 2718–2728

# Introduction



*fd*-virus



Isotropic

Nematic

Smectic

- Mono-disperse ( $L_{\text{length}}=880 \text{ nm}$ ,  $d_{\text{dia.}}=6.6 \text{ nm}$ ,  $P_L=2200 \text{ nm}$ )
- Negatively charged ( $10 \text{ e}^-/\text{nm}$  at  $8.2 \text{ pH}$ )
- Surface groups ( $-\text{COOH}$ ,  $-\text{OH}$ ,  $-\text{NH}_2$ )

[www.elsie.brandeis.edu/.../FD\\_Virus.htm](http://www.elsie.brandeis.edu/.../FD_Virus.htm)

<http://www.rowland.harvard.edu/rjf/dogic/fdvirus.php>

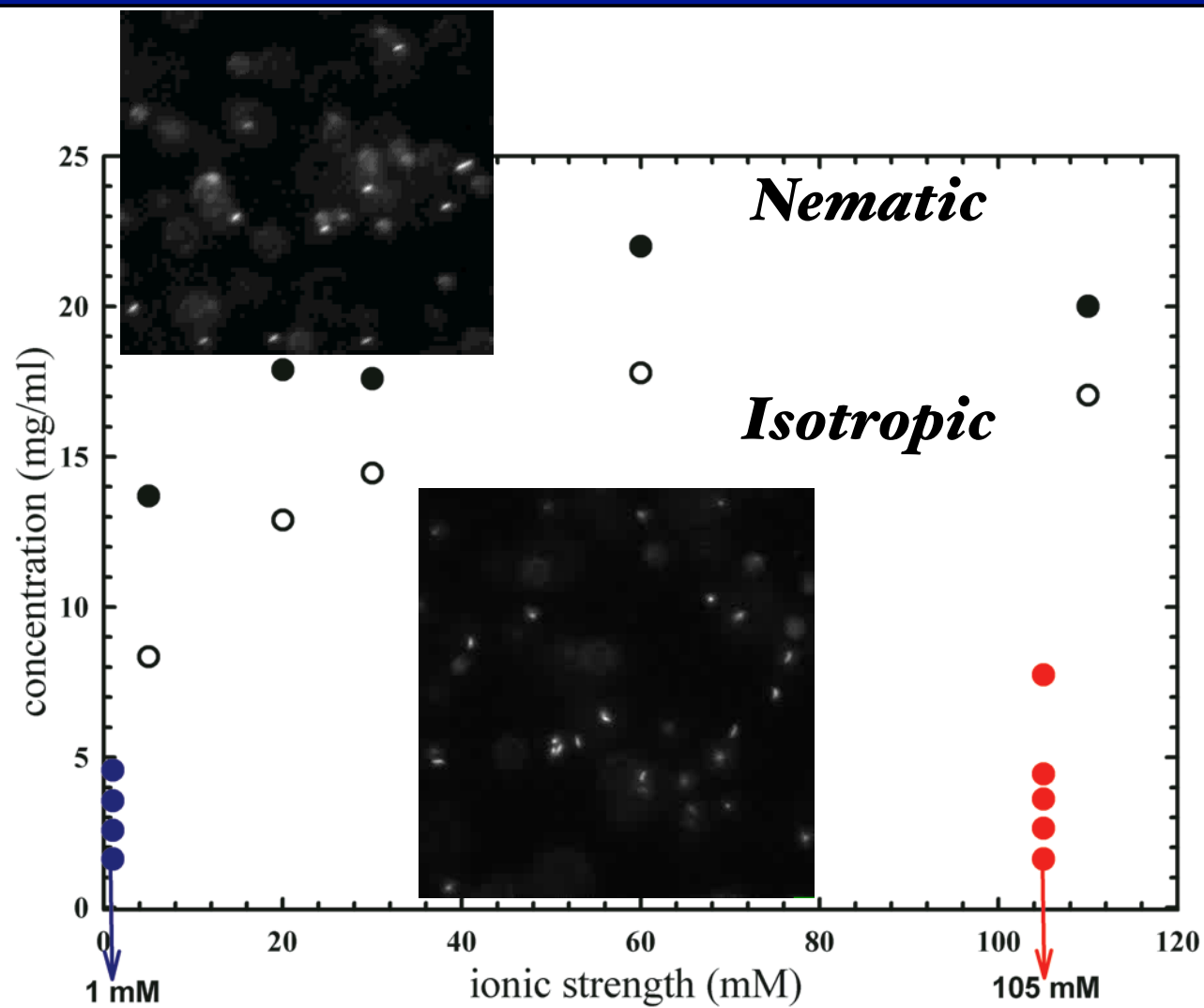
M. Paul Lettinga and Eric Grelet Phys. Rev. Lett. 99, 197802 (2007)



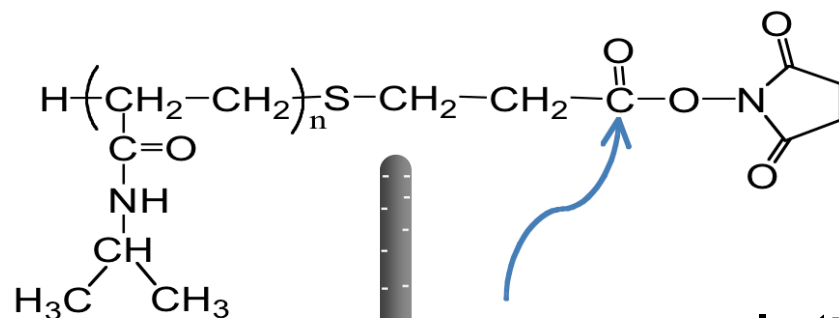
# Outline

1. Introduction
- 2. Phase diagram and gel structure**
3. Results
  - Gel point
  - Rheology
  - Scattering
4. Conclusions

# Phase diagram **solution**

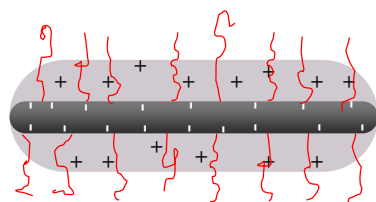
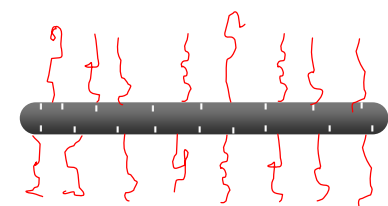


# Gelation

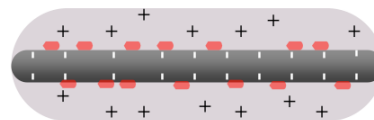


-(NH<sub>2</sub>)<sub>n</sub> poly(N-isopropylacrylamide)

32°C



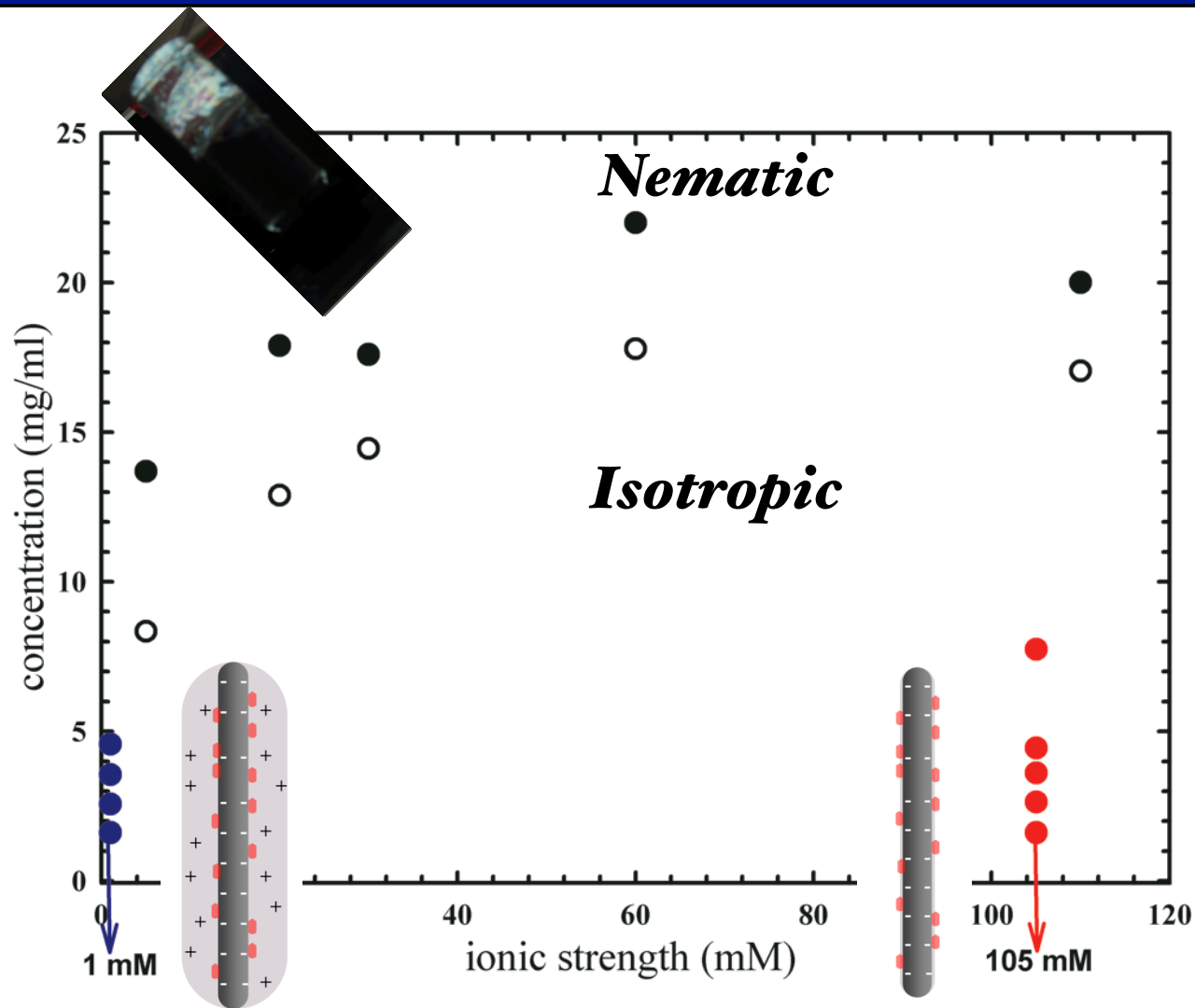
$T > T_g$



$$D_{eff}^{Polymer} = D_{fd} + 4R_g \quad 10 \text{ nm}$$

$$D_{eff}^{electrostatic} = D_{fd} + f(k^{-1}) \quad 26 \text{ nm}$$

# Phase diagram gel



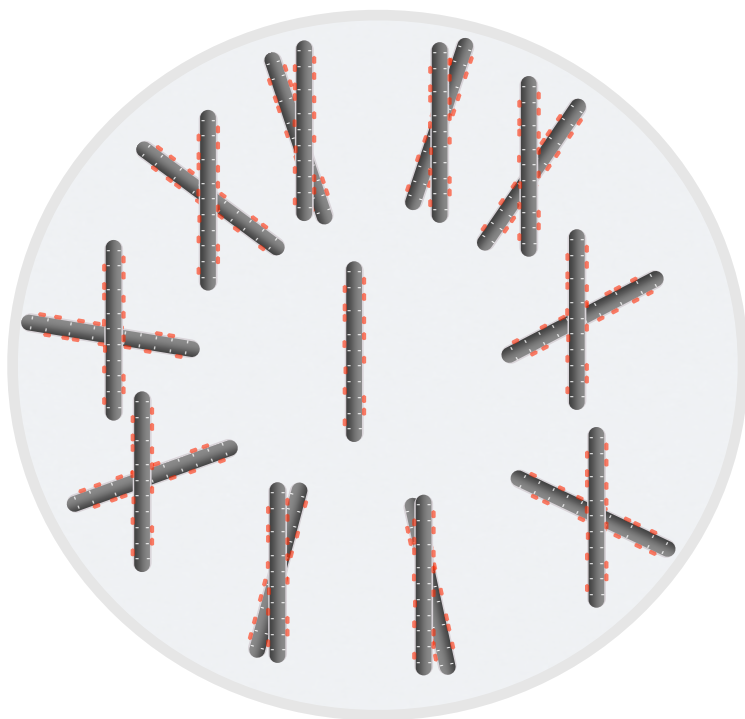
Zhang et al. Langmuir, 2009, 25 (4), 2437

# Possible gel structure

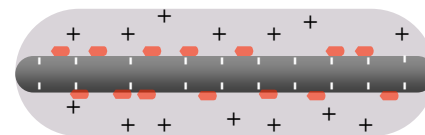
105 mM



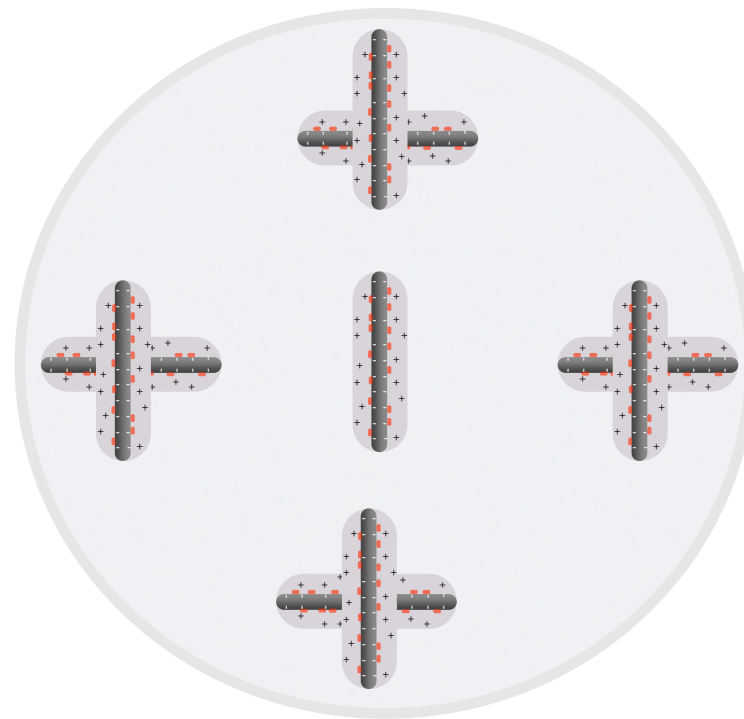
*Diffusion*



1 mM



*Diffusion* + *barrier*

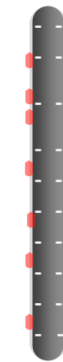
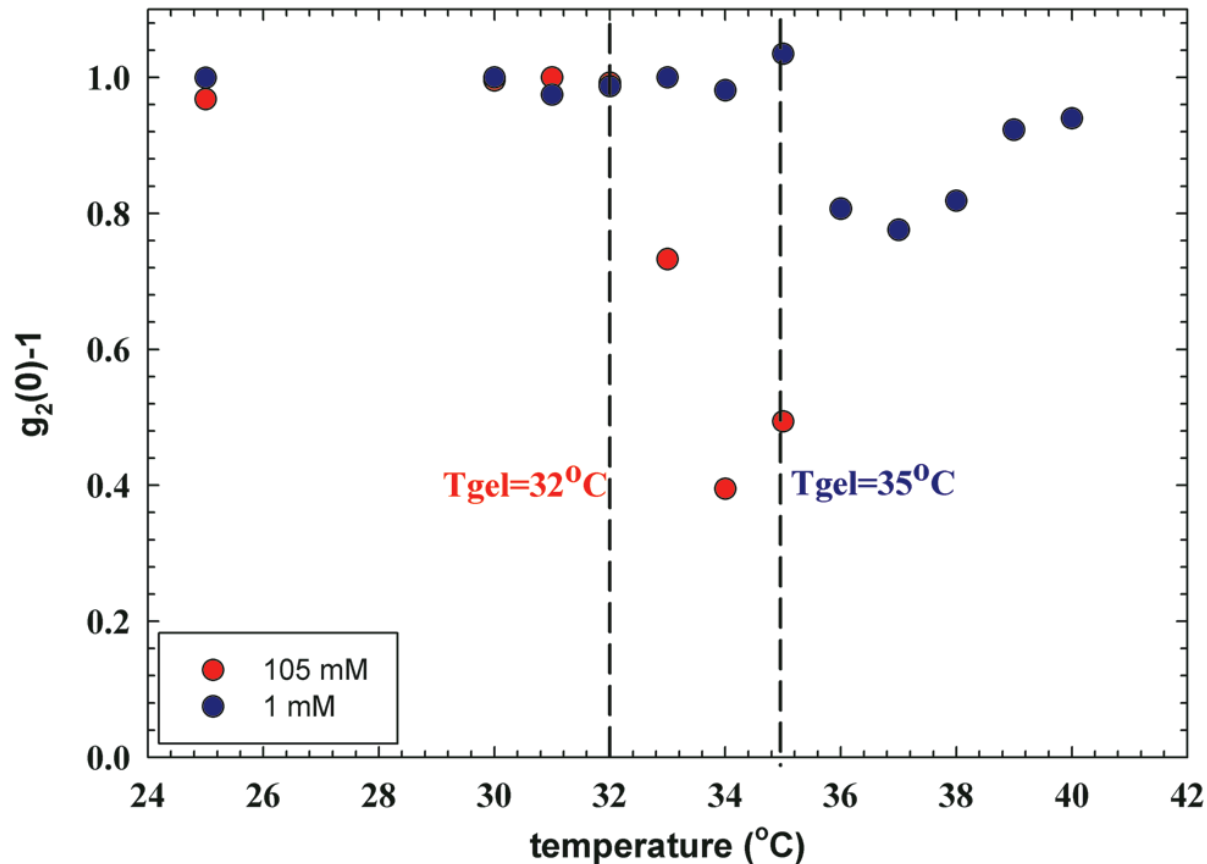




# Outline

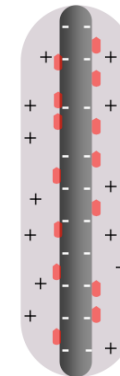
1. Introduction
2. Phase diagram and gel structure
3. Results
  - Gel point: *Dynamic light scattering*
  - Rheology
  - Scattering
4. Conclusions

# Gel point: Dynamic light scattering



105 mM

$T_{\text{gel}} = 32^{\circ}\text{C}$



1 mM

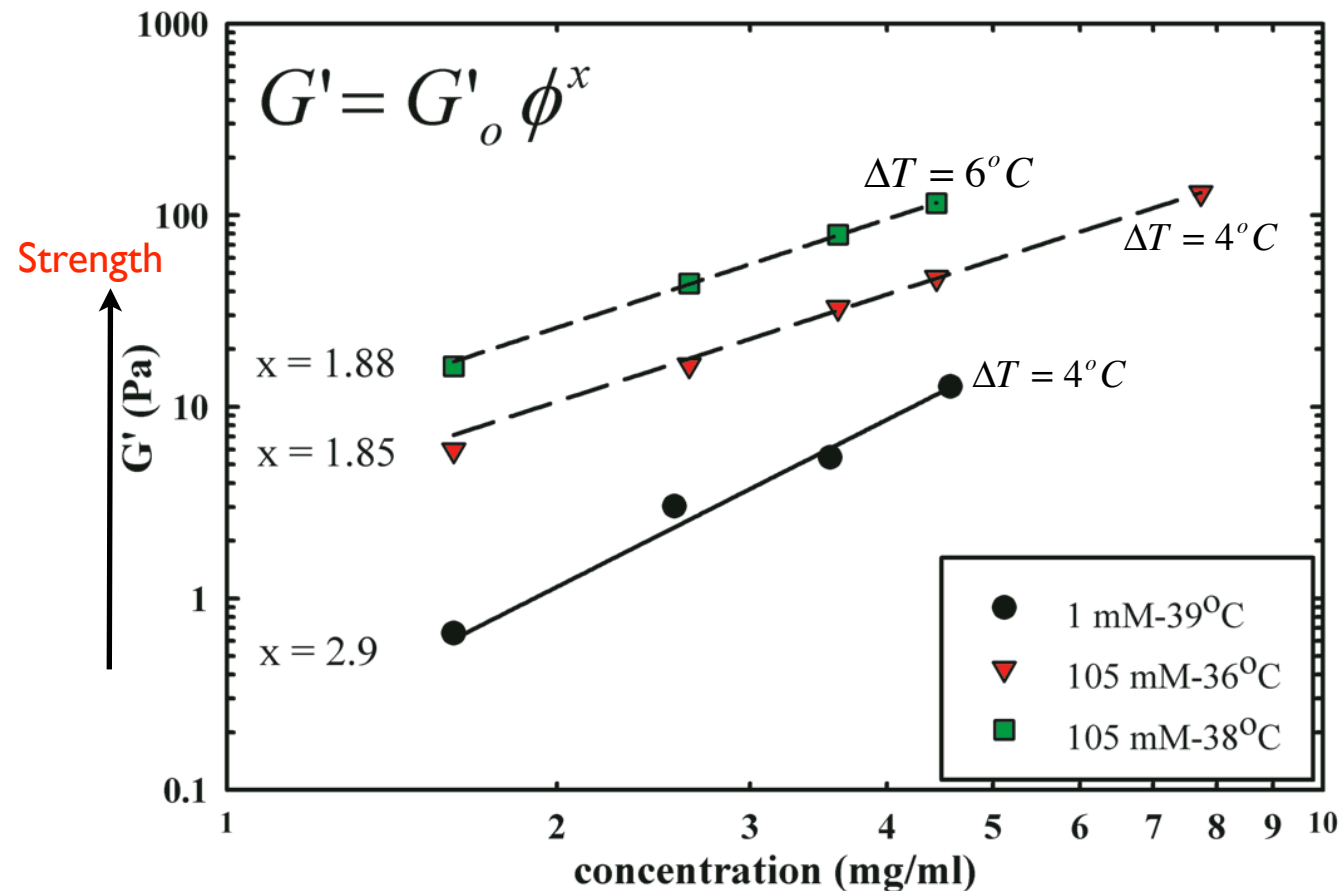
$T_{\text{gel}} = 35^{\circ}\text{C}$

*Shift in gelation temperature due to electrostatic double layer*

# Outline

1. Introduction
2. Phase diagram and gel structure
3. Results
  - Gel point
  - Rheology
    - Linear Viscoelasticity
      - *Strength and structure*
    - Yield stress
  - Scattering
4. Conclusions

# Storage modulus



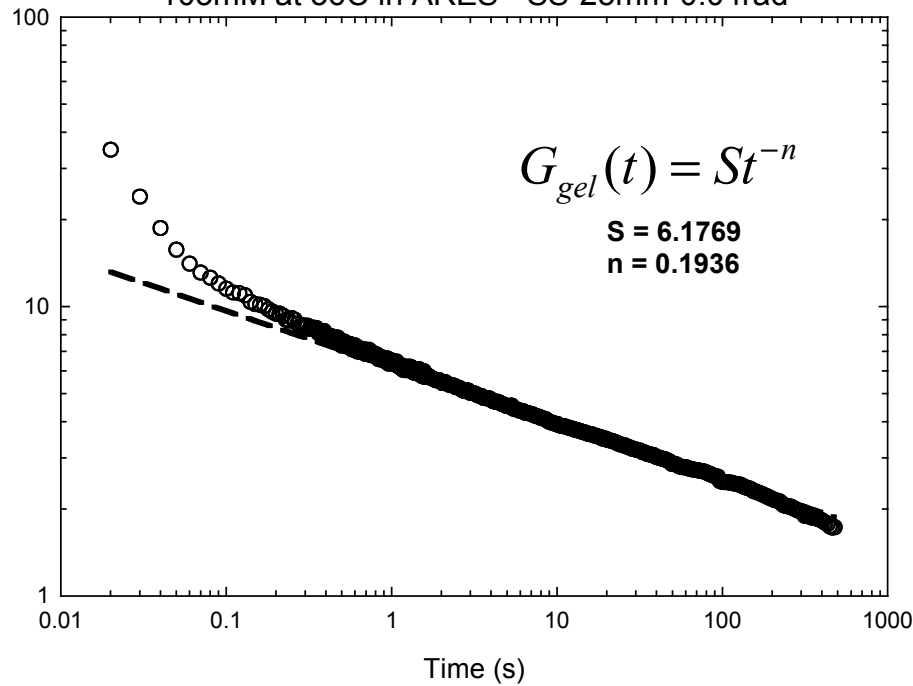
105 mM

1 mM

Characteristic length or structure changes with ionic strength but not with temperature

# Stress Relaxation

StressRelaxation for 1.835mg/ml of fd-Nipam in  
105mM at 36C in ARES - SS-25mm-0.04rad

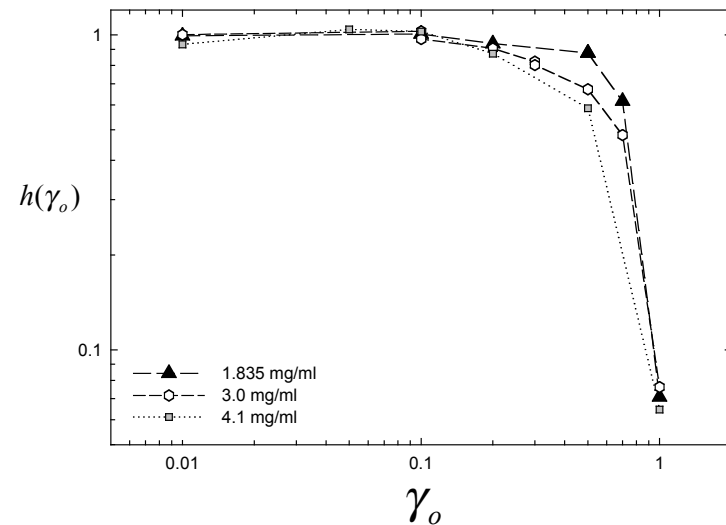


conc. (mg/ml) 105 mM	G'	
	S	n
1,61	7,05	0,149
2,64	18,9	0,15
3,61	36,0	0,15

105 mM

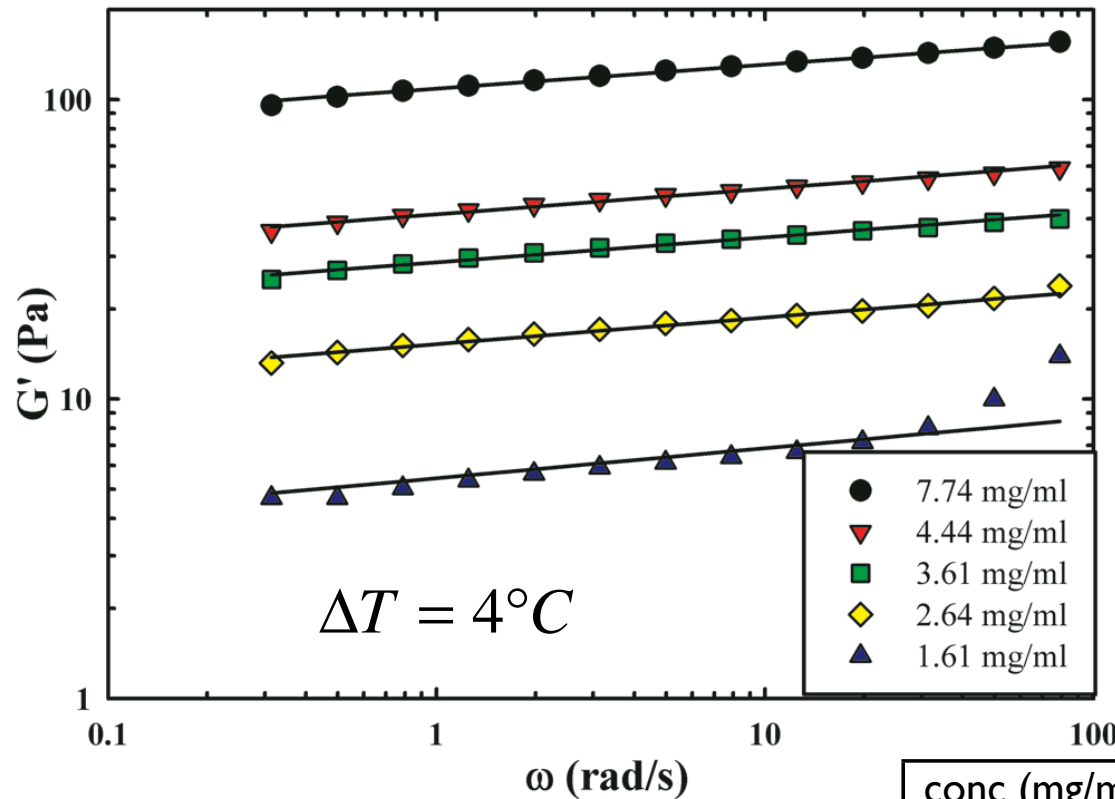


Normalized Intercept for the fit of the stress  
relaxation to  $G(t) = St^{-n}$





# Frequency sweep



105 mM

Critical gel

$$G(t) = St^{-n}$$

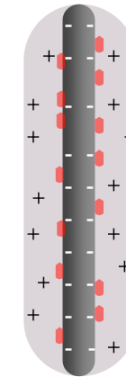
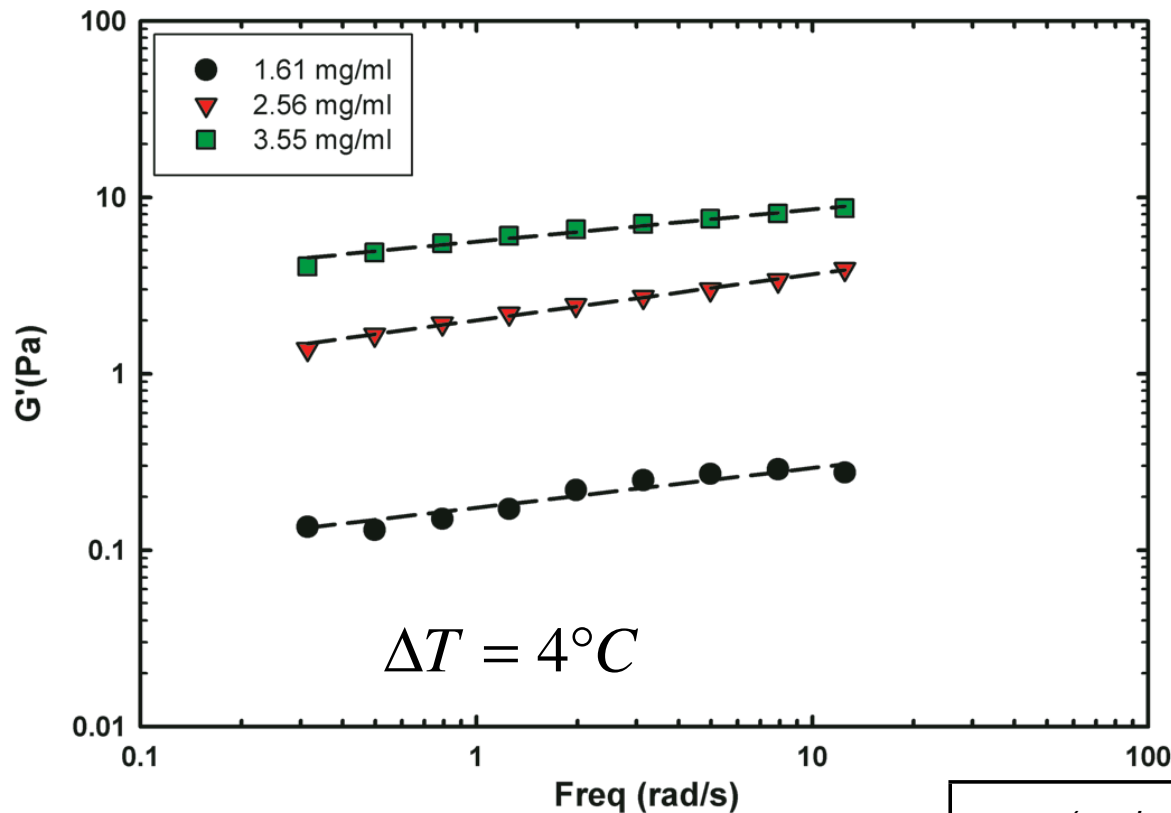
Strength      Structure

$$G'_{gel}(\omega) = \Gamma(1-n)\cos\left(\frac{n\pi}{2}\right)S\omega^n$$

Winter et al., JOR. 30, 367 (1986)

conc (mg/ml)	G'	
	S +/- 0.5	n +/- 0.002
105 mM		
1,61	7,0	0,12
2,64	17,0	0,12
3,61	32,0	0,12
4,44	46,0	0,12
7,74	120,0	0,12

# Frequency sweep



1 mM

Critical gel

$$G(t) = S t^{-n}$$

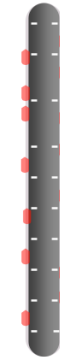
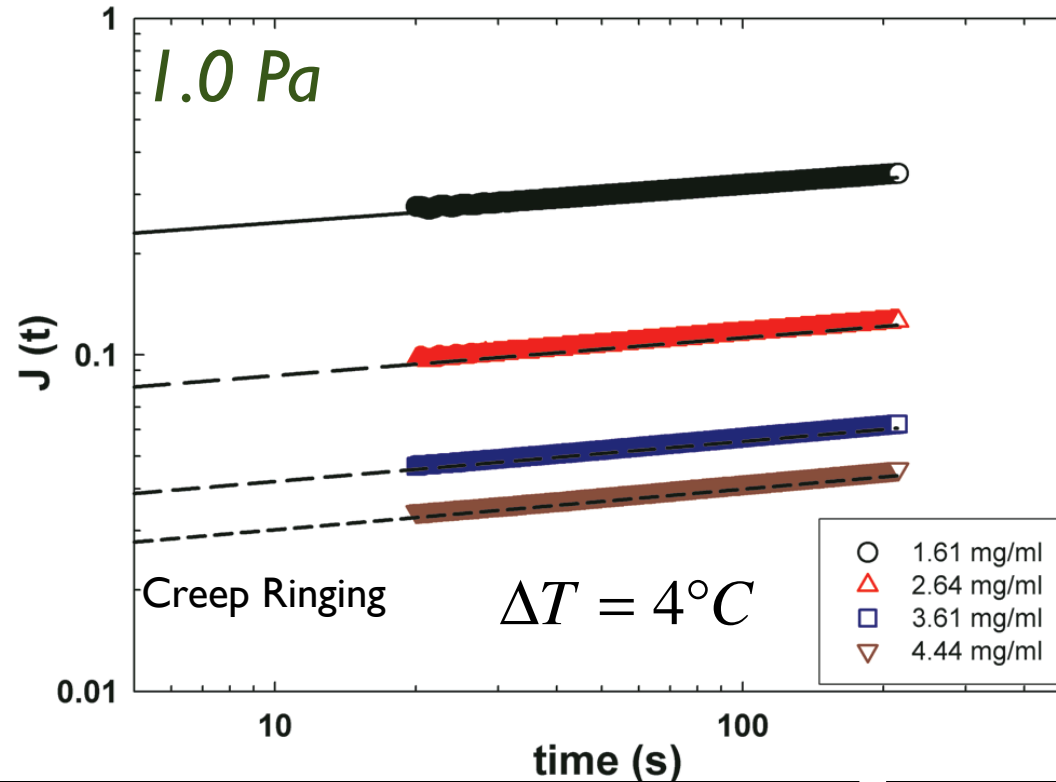
Strength      Structure

$$G'_{gel}(\omega) = \Gamma(1 - n) \cos\left(\frac{n\pi}{2}\right) S \omega^n$$

Winter et al., JOR. 30, 367 (1986)

conc (mg/ml) 1 mM	G'	
	S +/- 0.05	n +/- 0.04
1,61	0,15	0,22
2,54	1,7	0,22
3,55	5,0	0,22

# Creep



105 mM

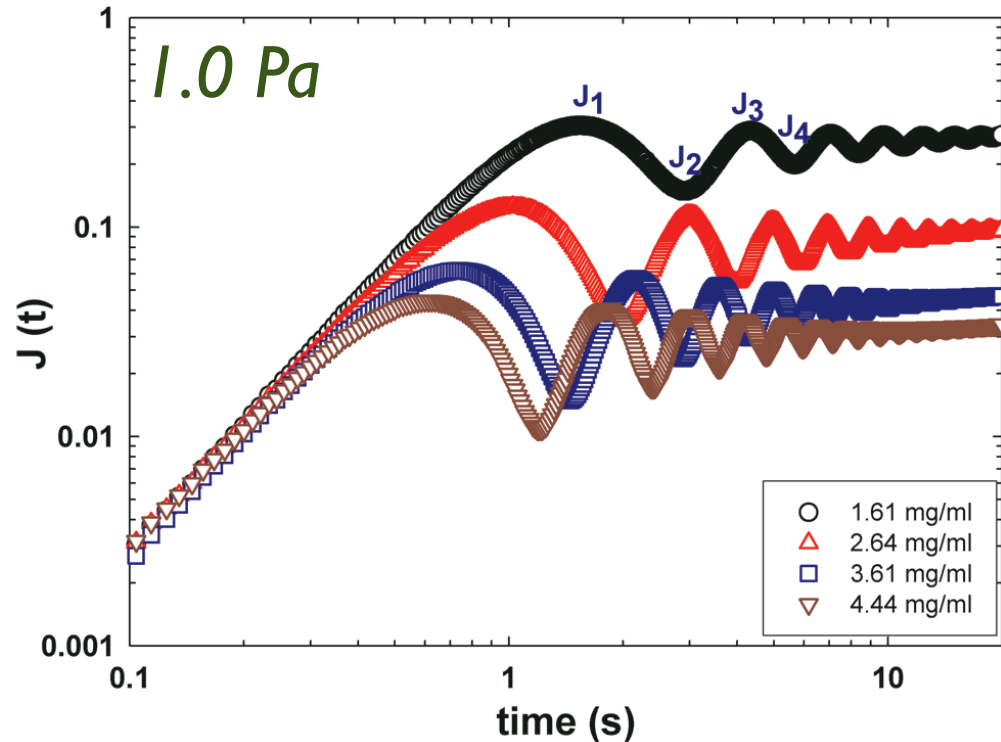
$$J(t) = \frac{t^n}{S\Gamma(1-n)\Gamma(1+n)}$$

Venkataraman et al. Rheol. Acta, 29, 423 (1990)

conc (mg/ml) 105 mM	Oscillatory	
	$S \pm 0.5$	$n \pm 0.02$
1,61	7	0,12
2,64	17	0,12
3,61	32	0,12
4,44	46	0,12

conc (mg/ml) 105 mM	Creep	
	$S \pm 0.5$	$n \pm 0.02$
1,61	5	0,14
2,64	14	0,14
3,61	30	0,14
4,44	40	0,14

# Creep Ringing



Conc. (mg/ml)	Freq. (rad/s)	Creep Ringing		Forced Oscillations	
		G' (Pa)	G'' (Pa)	G' (Pa)	G'' (Pa)
1,61	2,0	6,9	0,55	5,6	0,62
2,64	3,2	17	1,06	17	1,8
3,61	4,4	32,2	1,75	32,8	3,5
4,44	5,3	47	2,25	49,5	5,35

$$\tan(\delta) \approx \frac{\Delta}{\pi}$$

$$\Delta = 2 \ln \left( \frac{J_1 - 2J_2 + J_3}{-J_2 + 2J_3 - J_4} \right)$$

$$G' \approx \frac{I\omega^2}{b} [1 + (\Delta/2\pi)^2]$$

$$G'' \approx \frac{I\omega^2}{b} (\Delta/\pi)$$

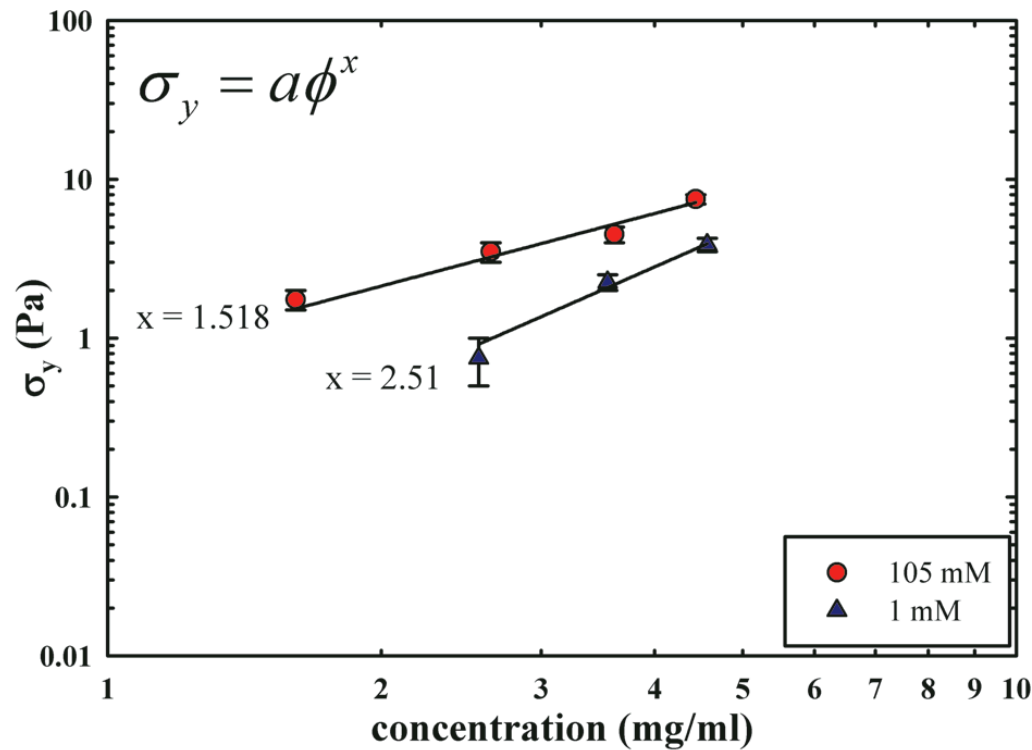
$$b_{c-p} = \frac{2\pi r^3}{3 \tan(\theta)}$$

Ewoldt et al. Rheol. Bull. 76, 4 (2007)



105 mM

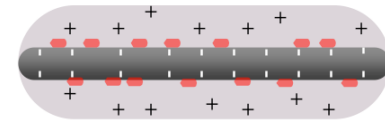
# Yield stress



105 mM



1 mM



$$x = \frac{d-1}{d-D_f}$$

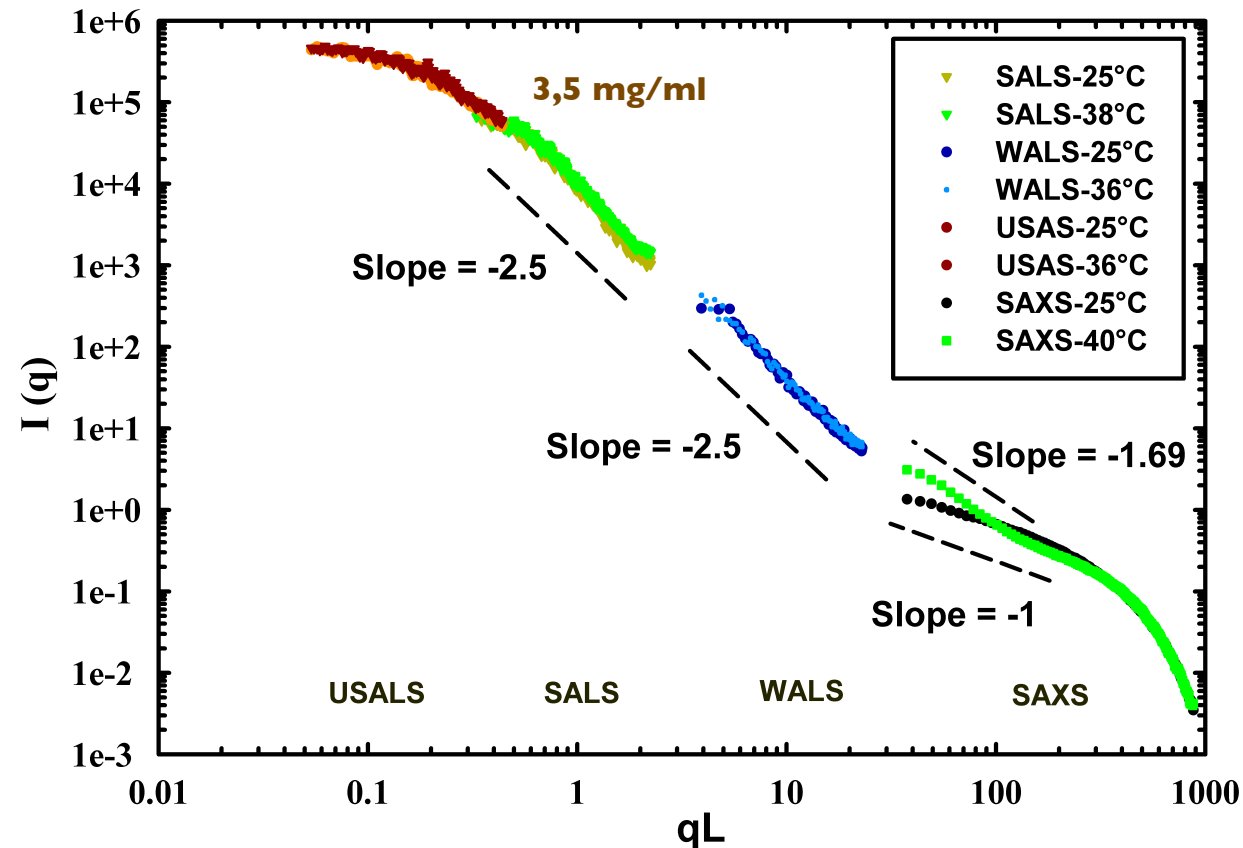
	<b>D<sub>f</sub></b> (rheology)	
<b>[I]</b>	<b>σ<sub>y</sub></b>	<b>G', γ<sub>c</sub></b>
<b>105 mM</b>	<b>1,7</b>	<b>1,7</b>
<b>1 mM</b>	<b>2,2</b>	<b>2,05</b>



# Outline

1. Introduction
2. Phase diagram and gel structure
- 3. Results**
  - Gel point
  - Linear Viscoelasticity
  - Yield stress
  - **Scattering**
4. Conclusions

# Scattering 105 mM



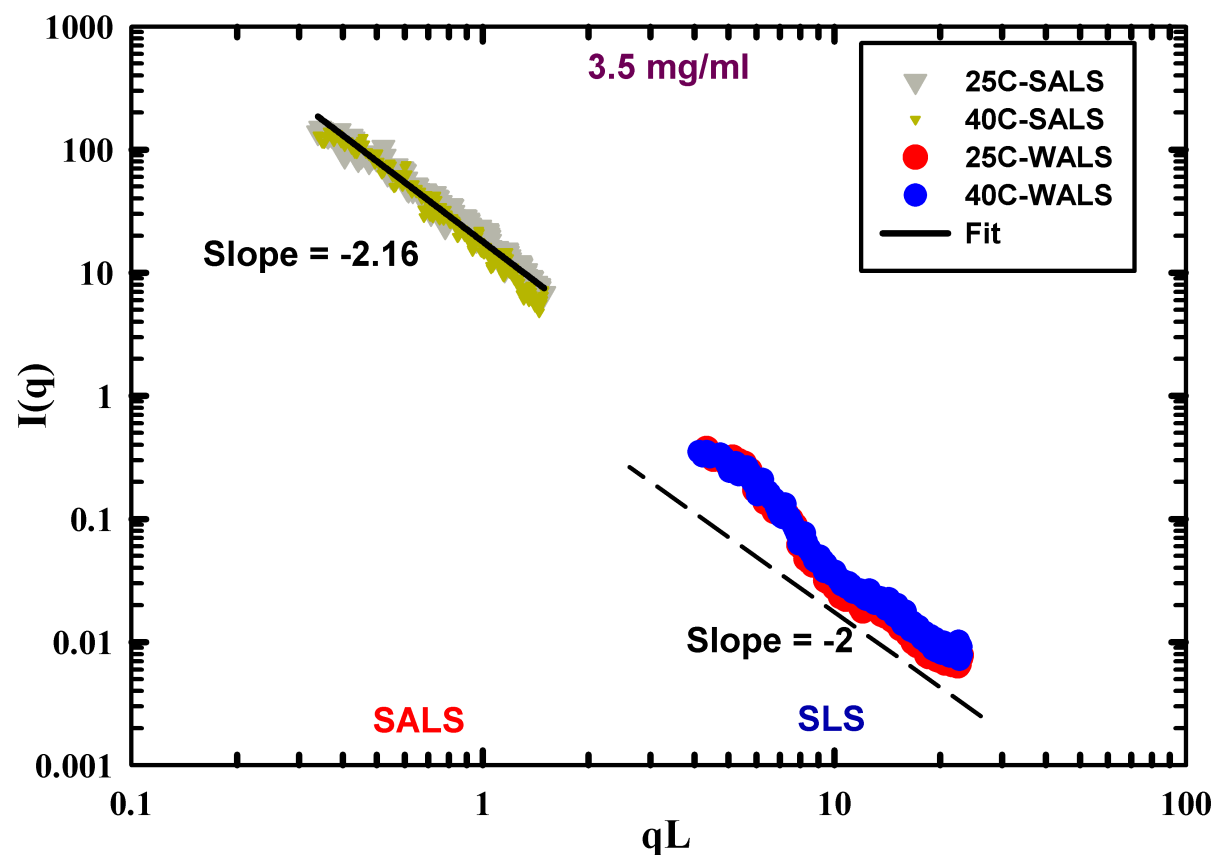
105 mM



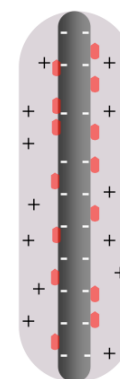
	$D_f$		
[I]	$\sigma_y$	$G', \gamma_c$	Scattering
105 mM	1,7	1,7	1,69*
1 mM	2,2	2,05	

\* low q in SAXS

# Scattering 1 mM



1 mM



\* low q in SAXS

	$D_f$		
[I]	$\sigma_y$	$G', \gamma_c$	Scattering
105 mM	1,7	1,7	1,69*
1 mM	2,2	2,05	2,16

# Conclusion

## 1. Gel structure and $T_{gel}$

- *ionic strength*

## 2. Linear viscoelastic limit

- *power law scaling - critical gel*
- *unique  $S(\text{conc. and } [I])$  and  $n([I])$*

## 3. Fractal dimension - yield stress and $G'$ & $\gamma_c$

- *1,7 for gels in 105 mM  $[I]$  (DLCA)*
- *2,2 for gels in 1 mM  $[I]$  (RLCA)*

## 4. Fractal dimension - scattering technique

- *1,7 for gels in 105 mM  $[I]$  (DLCA) - only at low  $q$  regime SAXS*
- *2,2 for gels in 1 mM  $[I]$  (RLCA)*



NANODIRECT

Thank you

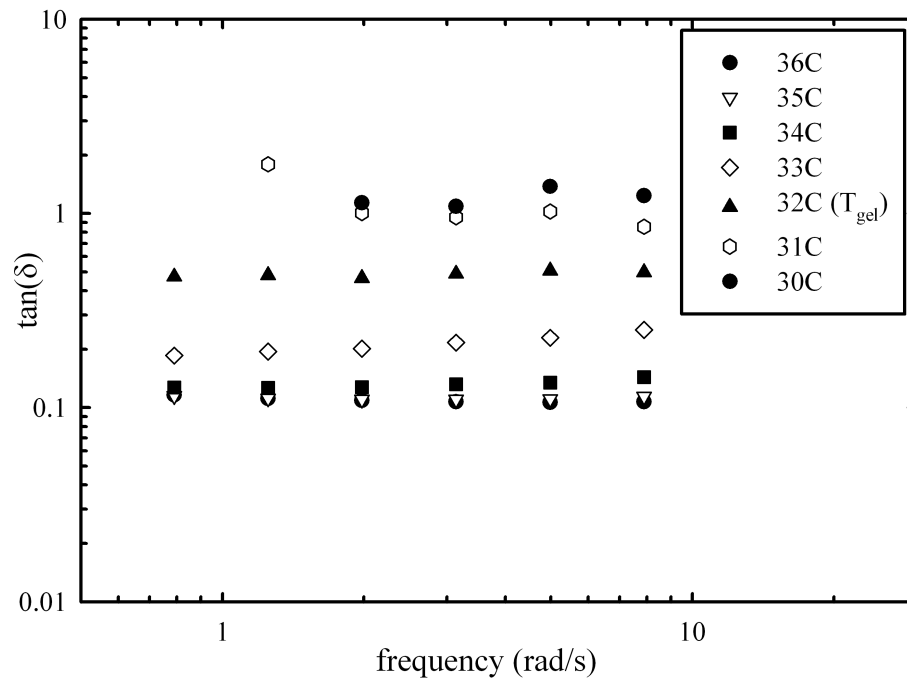


# No. grafted Polymer

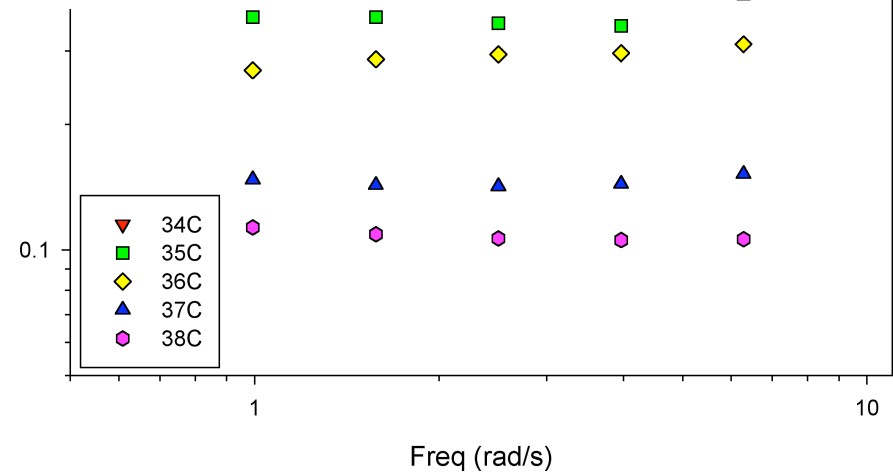
***Number of PNIPAM polymers per virus*** Since the refractive index increment ( $dn/dc$ ) is proportional to the mass density, the  $dn/dc$  difference between the bare virus and the PNIPAM-*fd* is due to the grafted polymer. This can be used to estimate the number of grafted polymers on each virus.<sup>10</sup> The  $dn/dc$  of carboxyl-terminated PNIPAM ( $(dn/dc)_p$ ) and *fd* virus ( $(dn/dc)_{fd}$ ) were determined with the homemade scanning Michelson differential interferometer at  $T = 25^\circ\text{C}$ .<sup>6</sup> As for PNIPAM-*fd* system, the concentration of *fd* virus in PNIPAM-*fd* suspensions can be easily obtained by UV absorption (PNIPAM does not absorb in the UV range where *fd* absorbs). So an apparent  $dn/dc$  can also be determined based on the virus concentration ( $(dn/dc)_{app}$ ). The number of PNIPAM grafted on each virus,  $N$ , can be estimated by the following equation<sup>1</sup>

$$\text{Vol}_{\text{eff}} = C_{fd} N_A L \pi (D_{\text{eff}} / 2)^2 / M_w$$

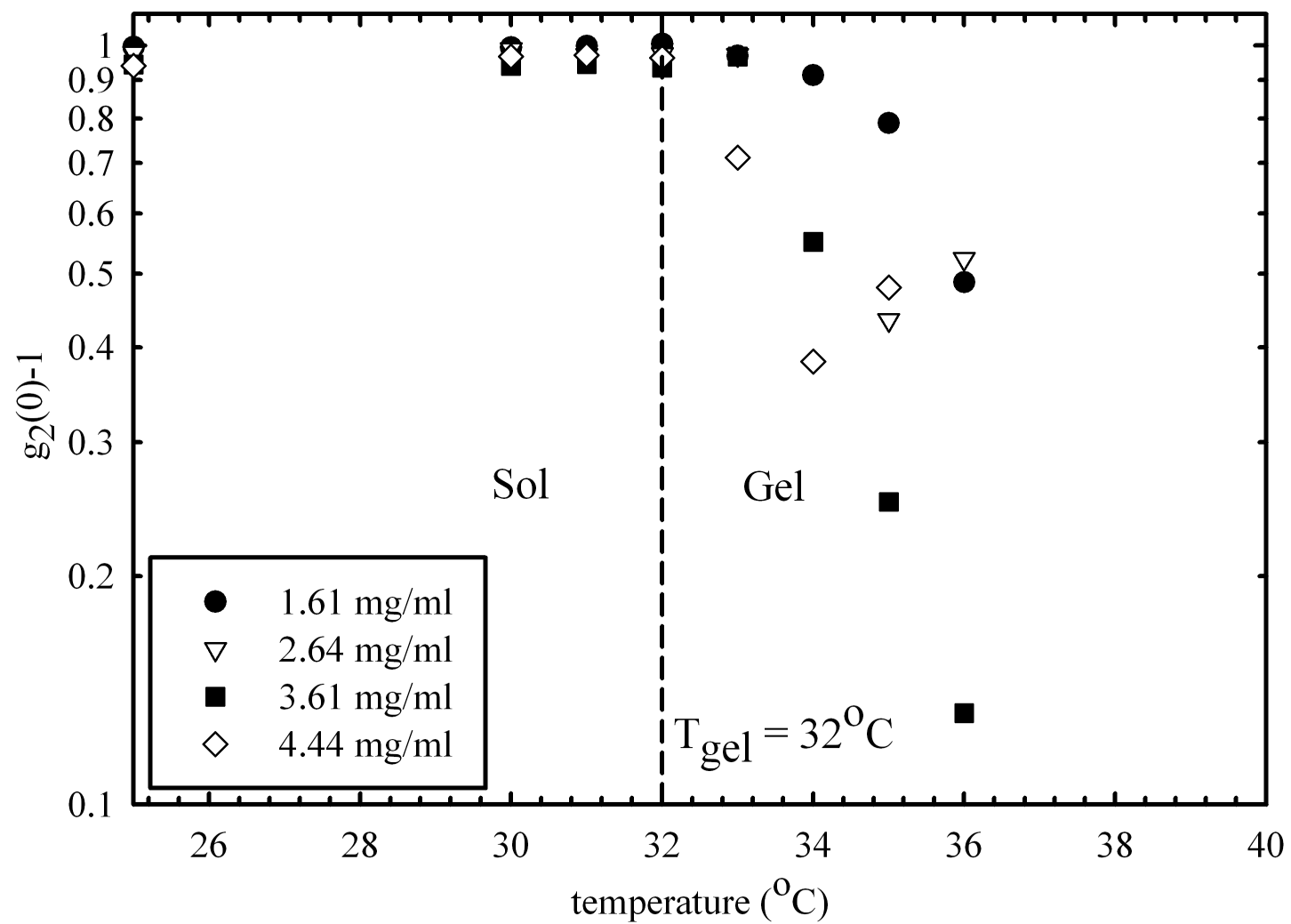
# Gel temperature from Rheology



4.1mg/ml in 5mM (38-33C) 5% strain (MCR)

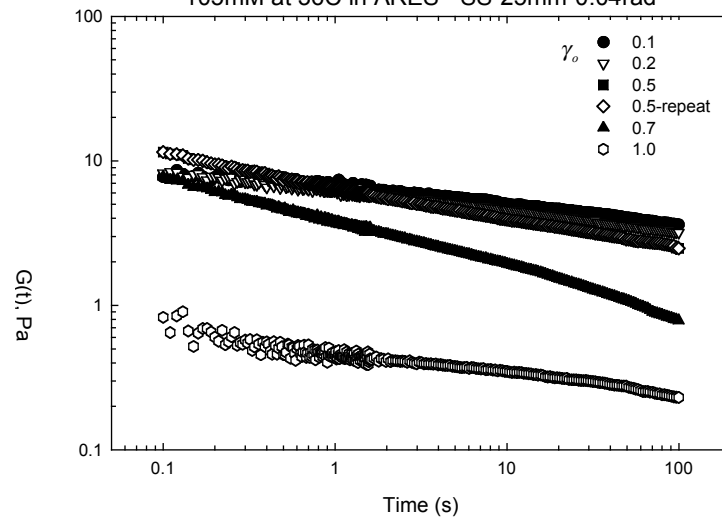


# DLS



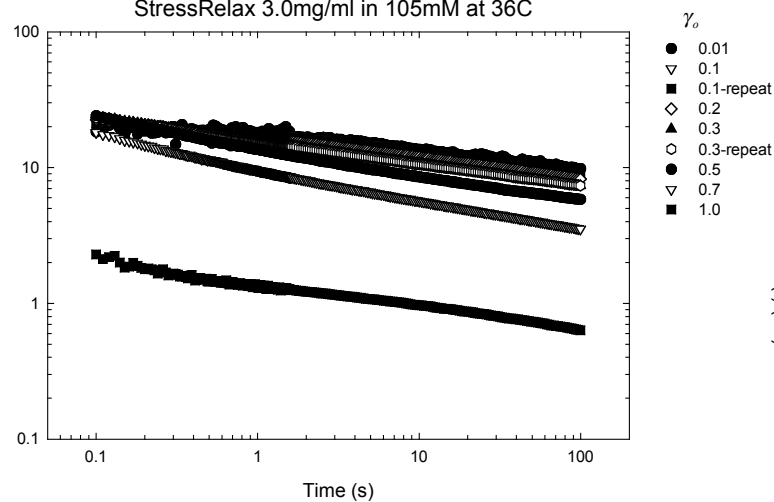
# Stress Relaxation

StressRelaxation for 1.835mg/ml of fd-Nipam in 105mM at 36C in ARES - SS-25mm-0.04rad

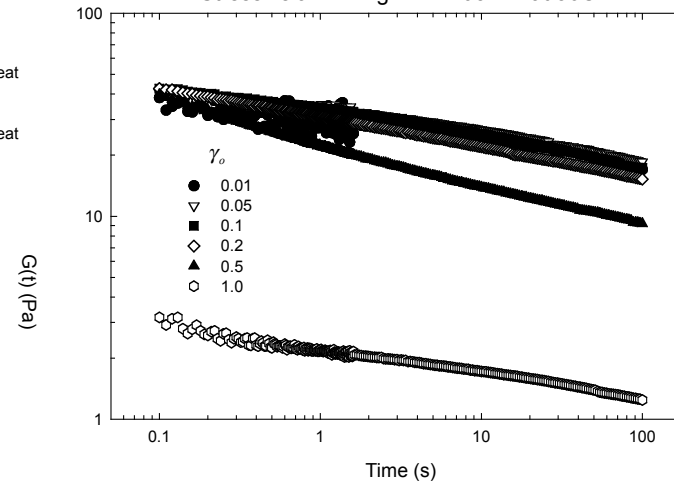


The yielding (nonlinear regime) strain decreases with the conc. (Meaning that the system become more brittle with inc. in conc.). Also can be seen in strain sweep (indicating brittle)

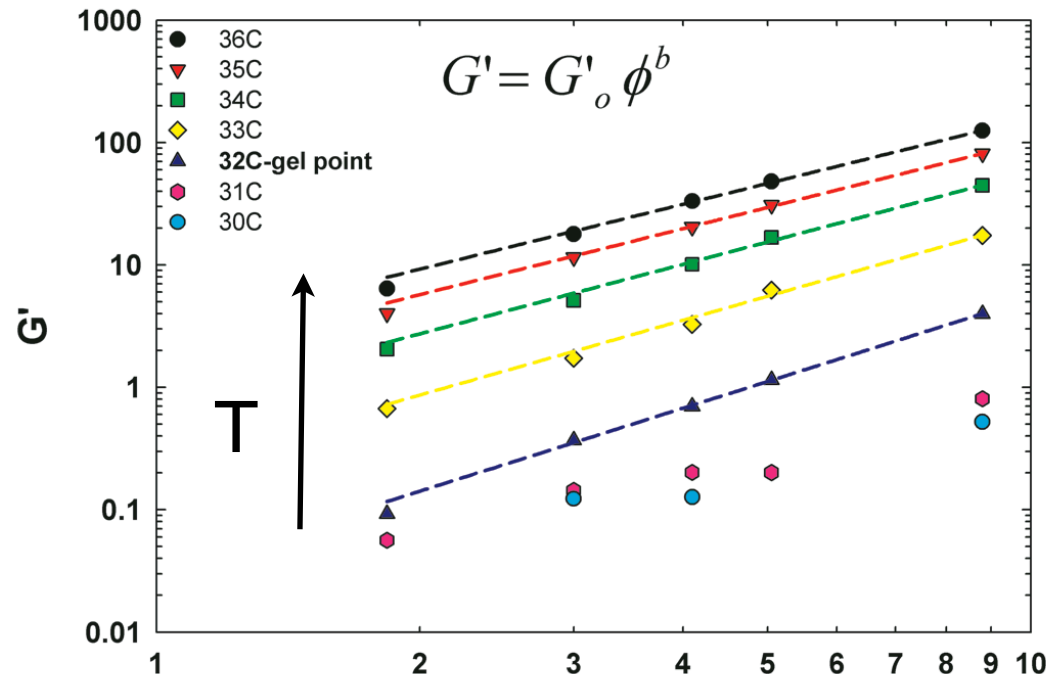
StressRelax 3.0mg/ml in 105mM at 36C



StressRelax 4.1mg/ml in 105mM at 36C



# Percolation theory scaling



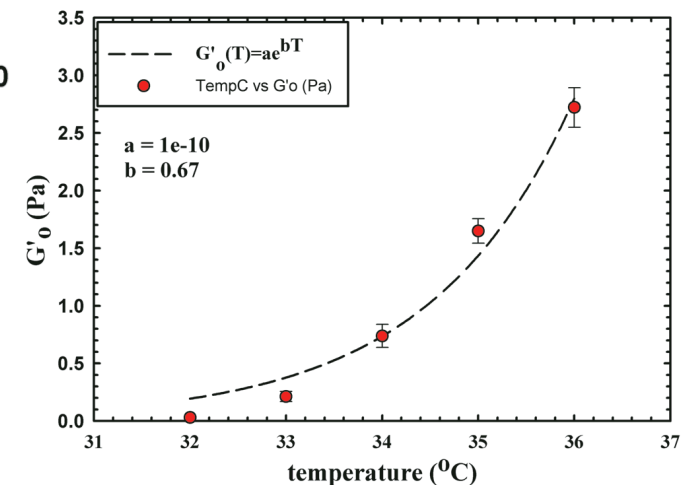
Temp	$G'_o$ (Pa)	b
36	2,72	1,76
35	1,65	1,79
34	0,74	1,88
33	0,21	2,03
32	0,03	2,25

$$G' = G_o (\phi - \phi_g)^b \quad \text{Grant and Russel}$$

$$G' = G_o \left( \frac{\phi}{\phi_g} - 1 \right)^b \quad \text{Zukoski et al.}$$

$$G' = \frac{G_o}{\phi_g^b} (\phi - \phi_g)^b = G_o(T) (\phi - \phi_g)^b$$

*Bergstrom et al.*

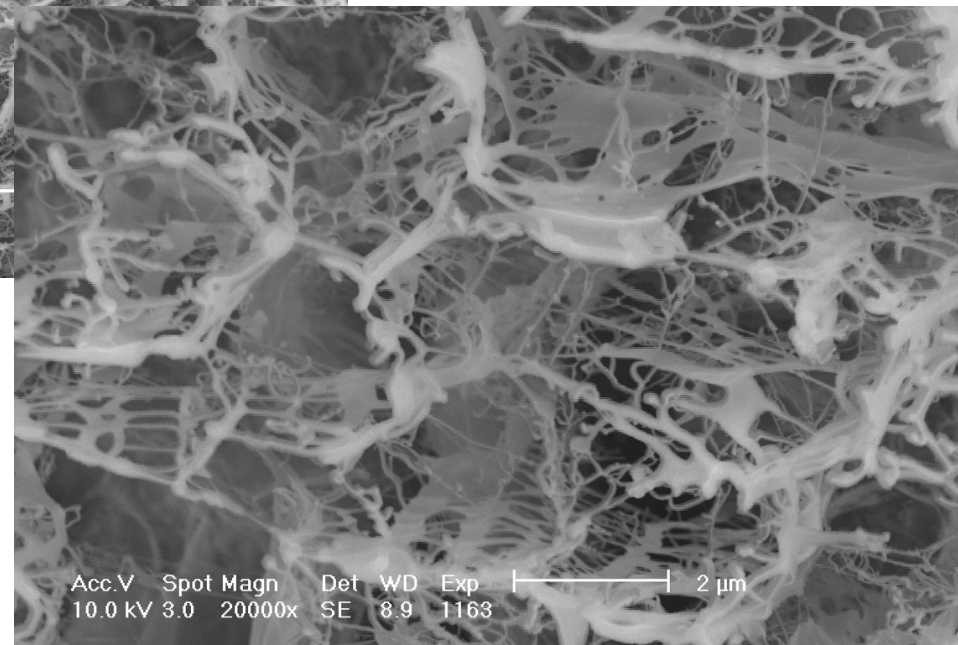
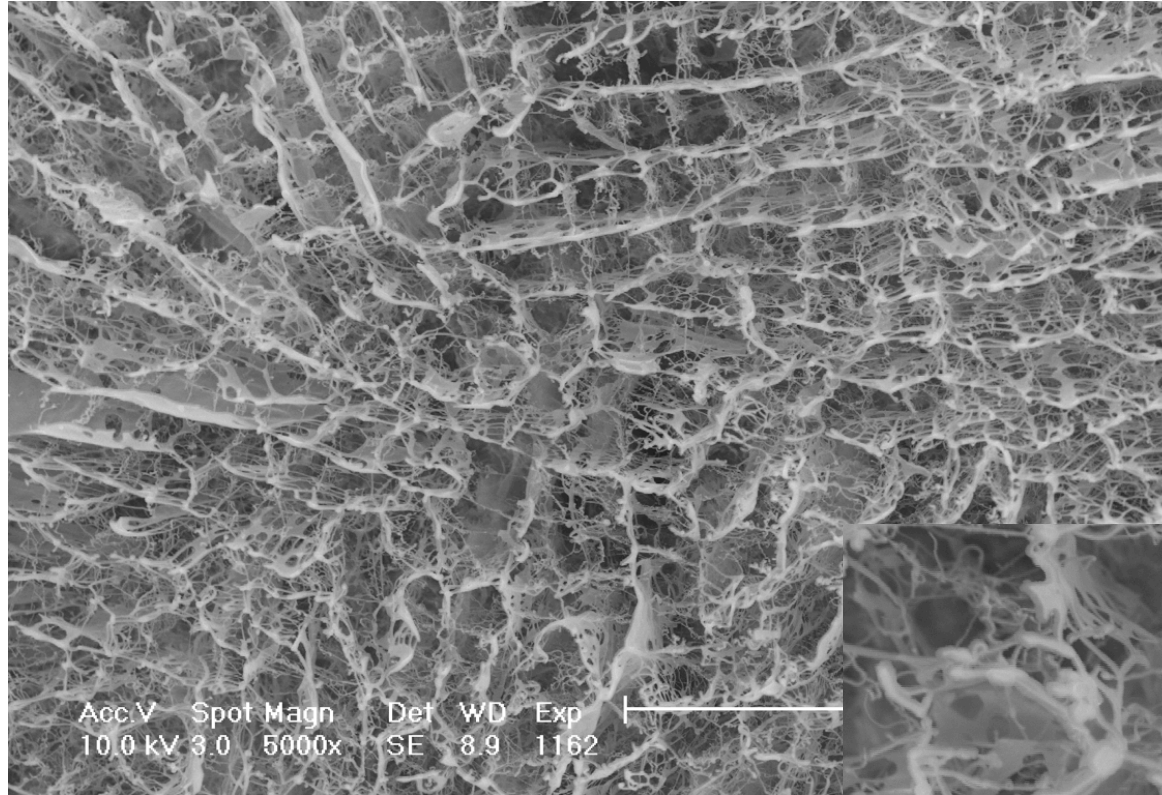


M. C. Grant and W. B. Russel, Phys. Rev. E 47, 2606 - 2614 (1993)

C. J. Rueb and C. F. Zukoski, Rheol. Volume 41, Issue 2, pp. 197-218 (March 1997)

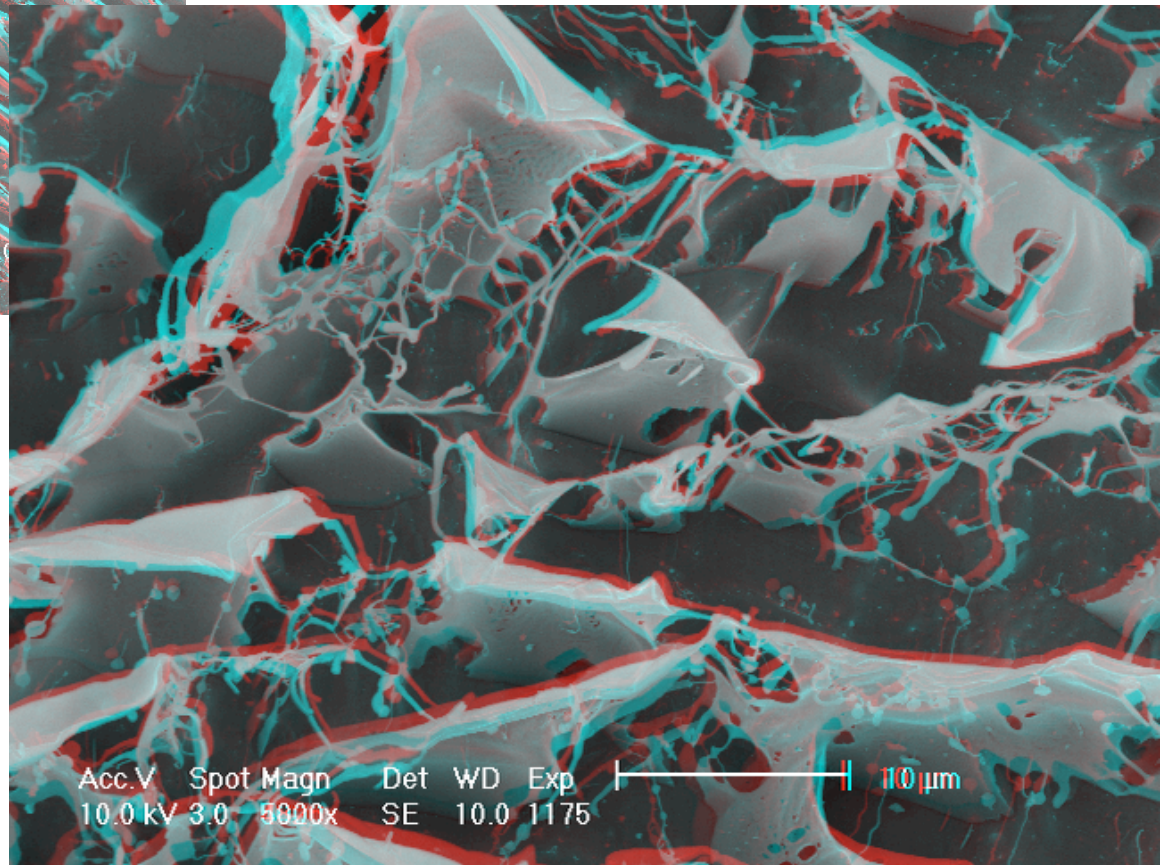
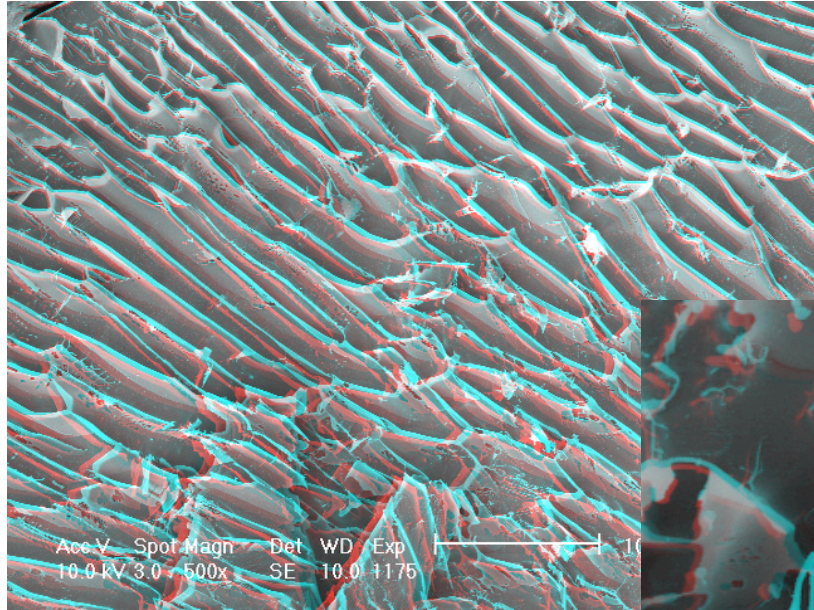
Yanez et al. J. Coll. Interf. Sci., 209, 162 (1999)

# Cryo SEM (1 mM) Liquid



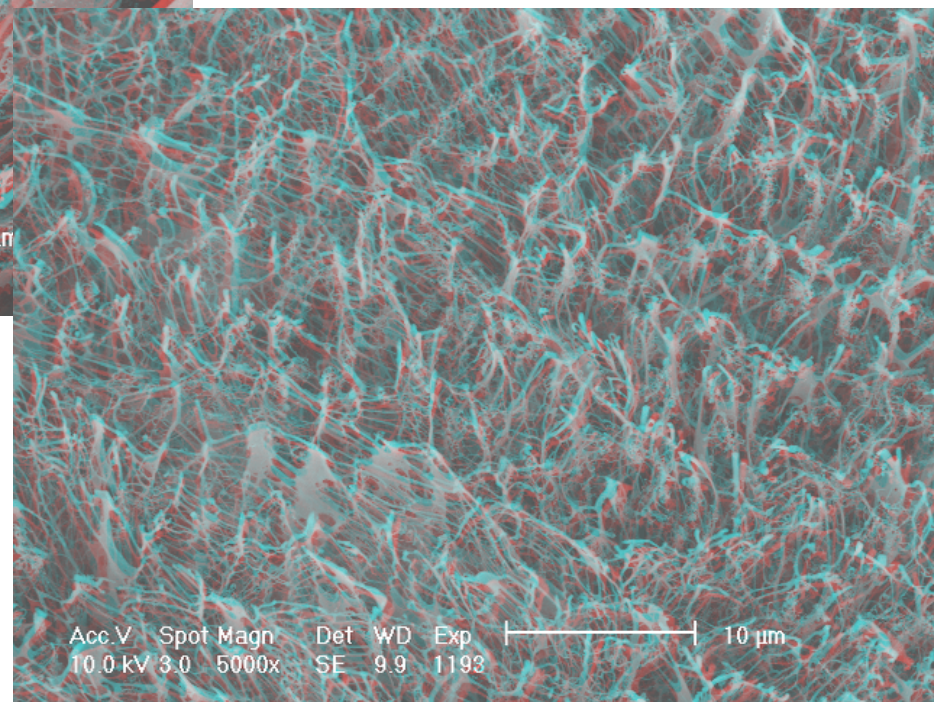
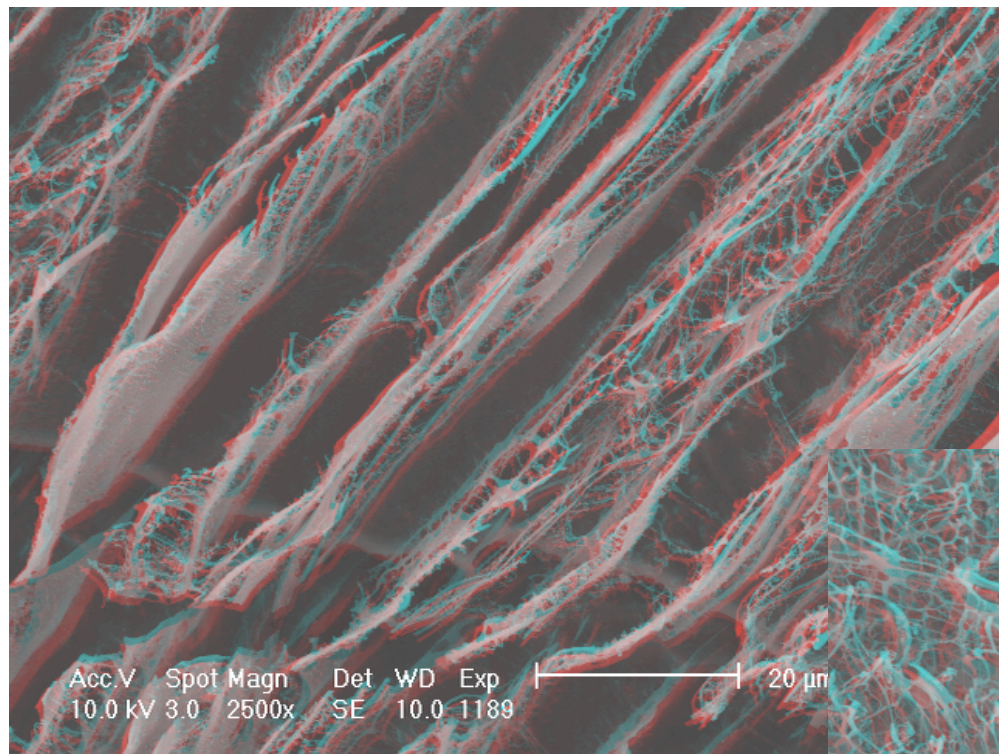


# Cryo SEM (105 mM) Liquid



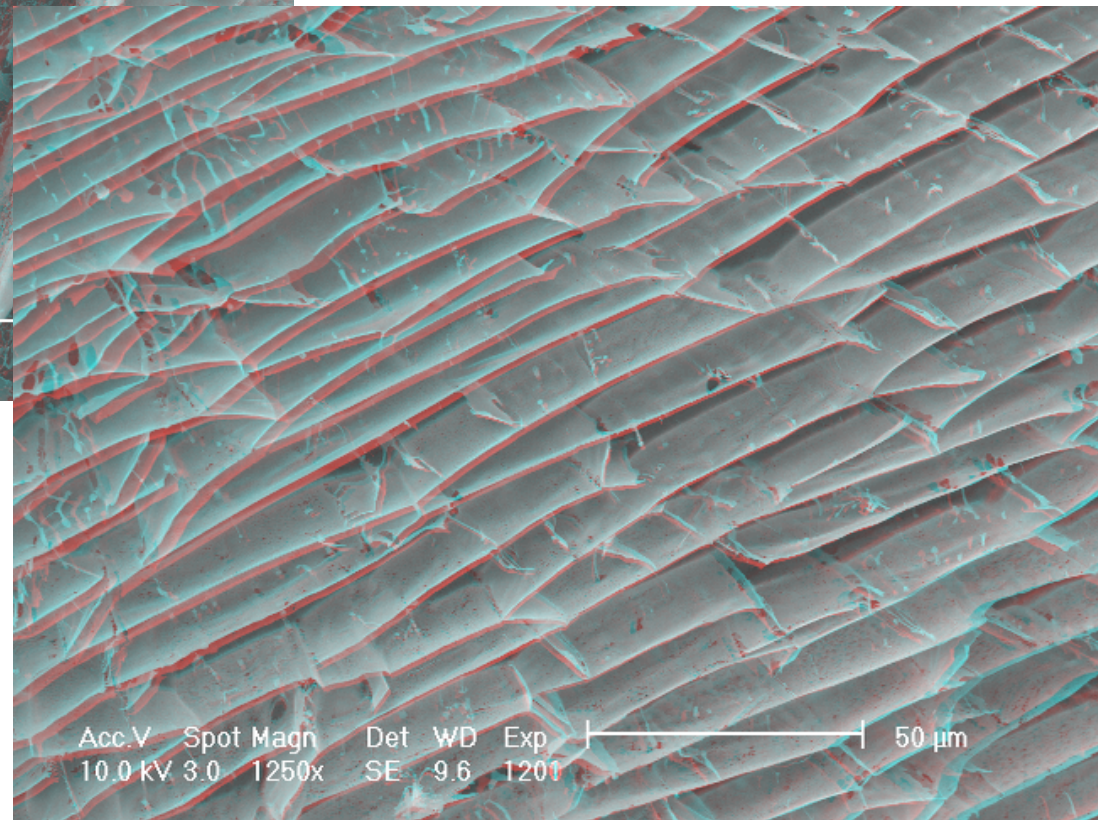
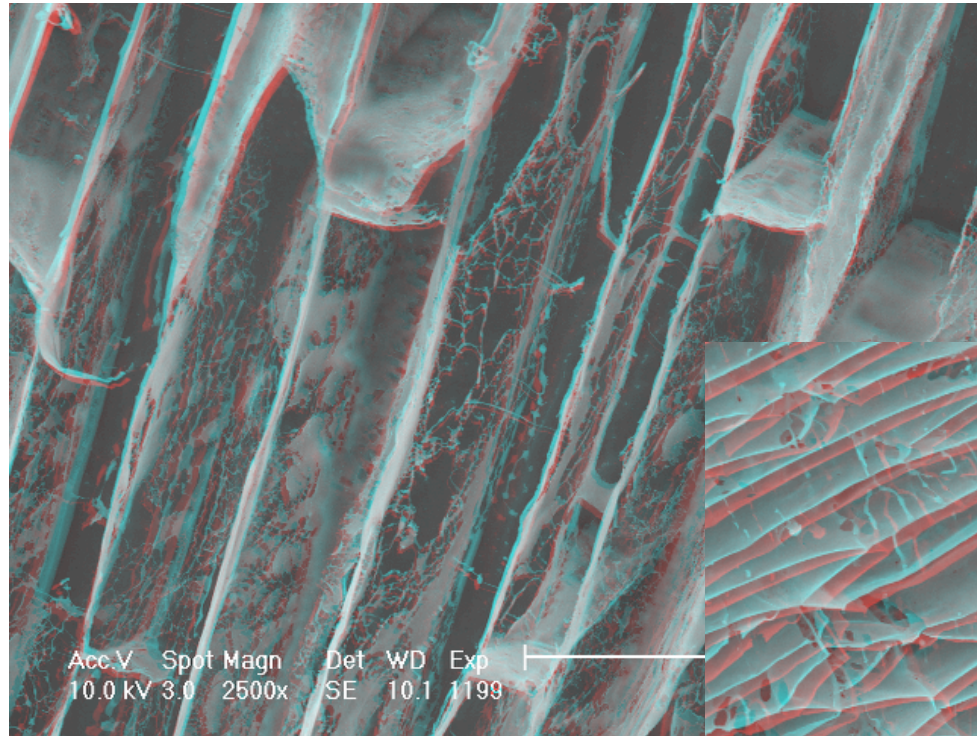


# Cryo SEM (1 mM) Gel



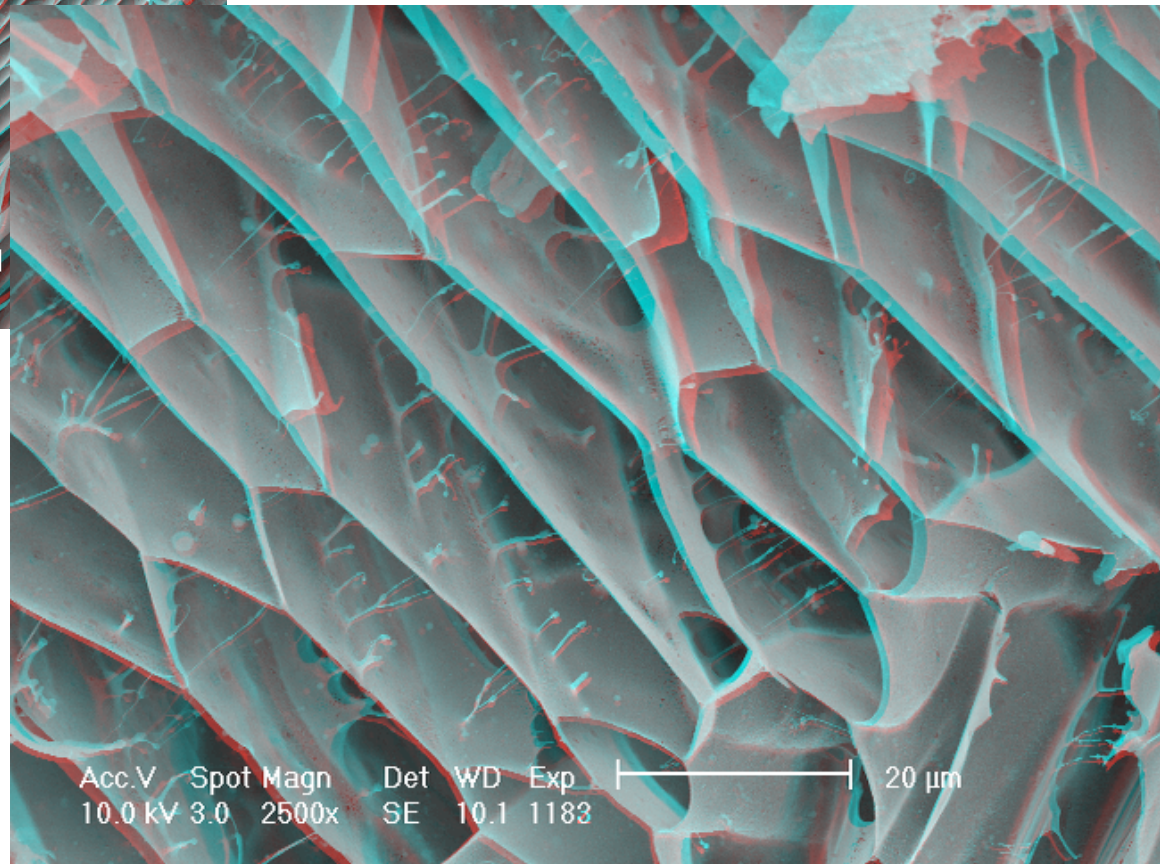
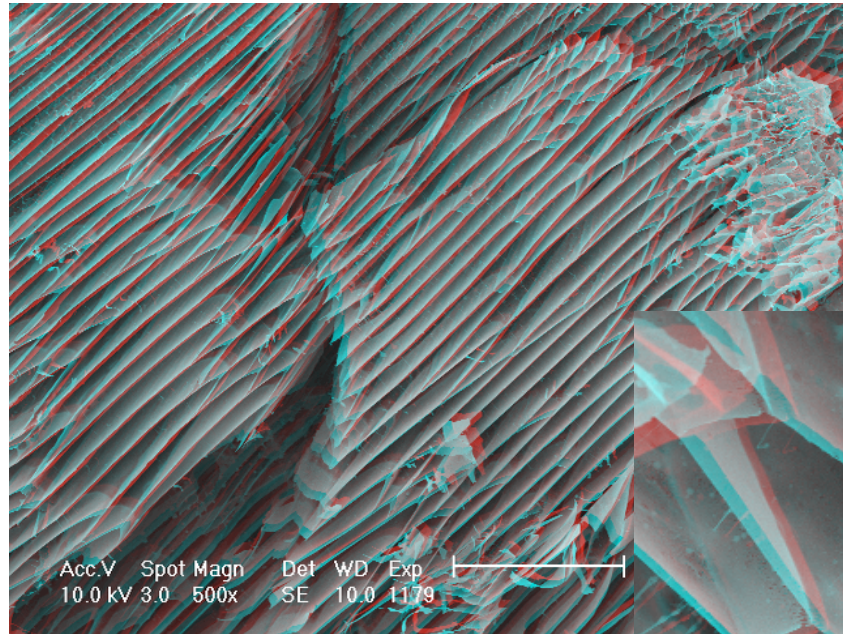


# Cryo SEM (105 mM) Gel

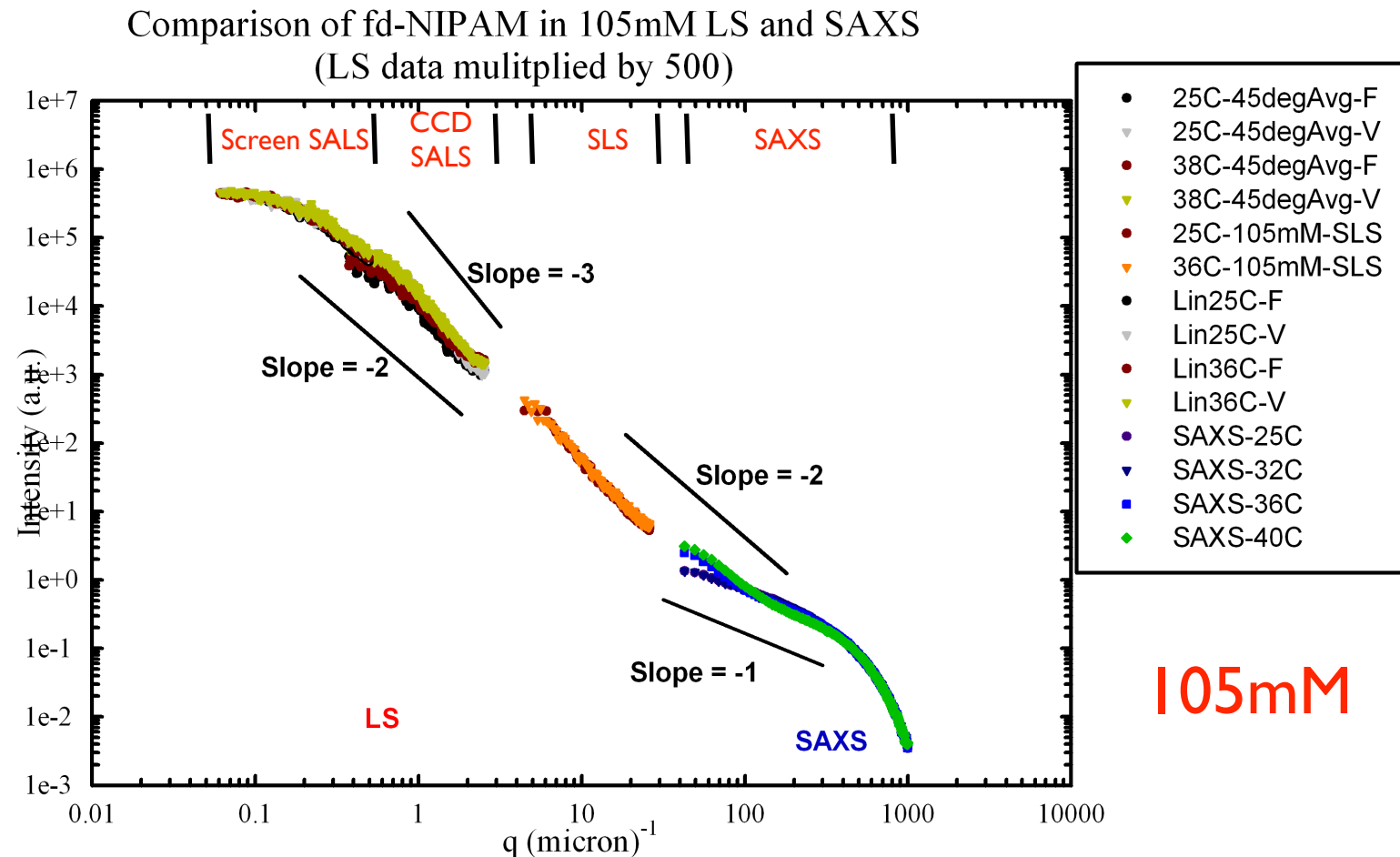




# Cryo SEM (Pf1 liquid)



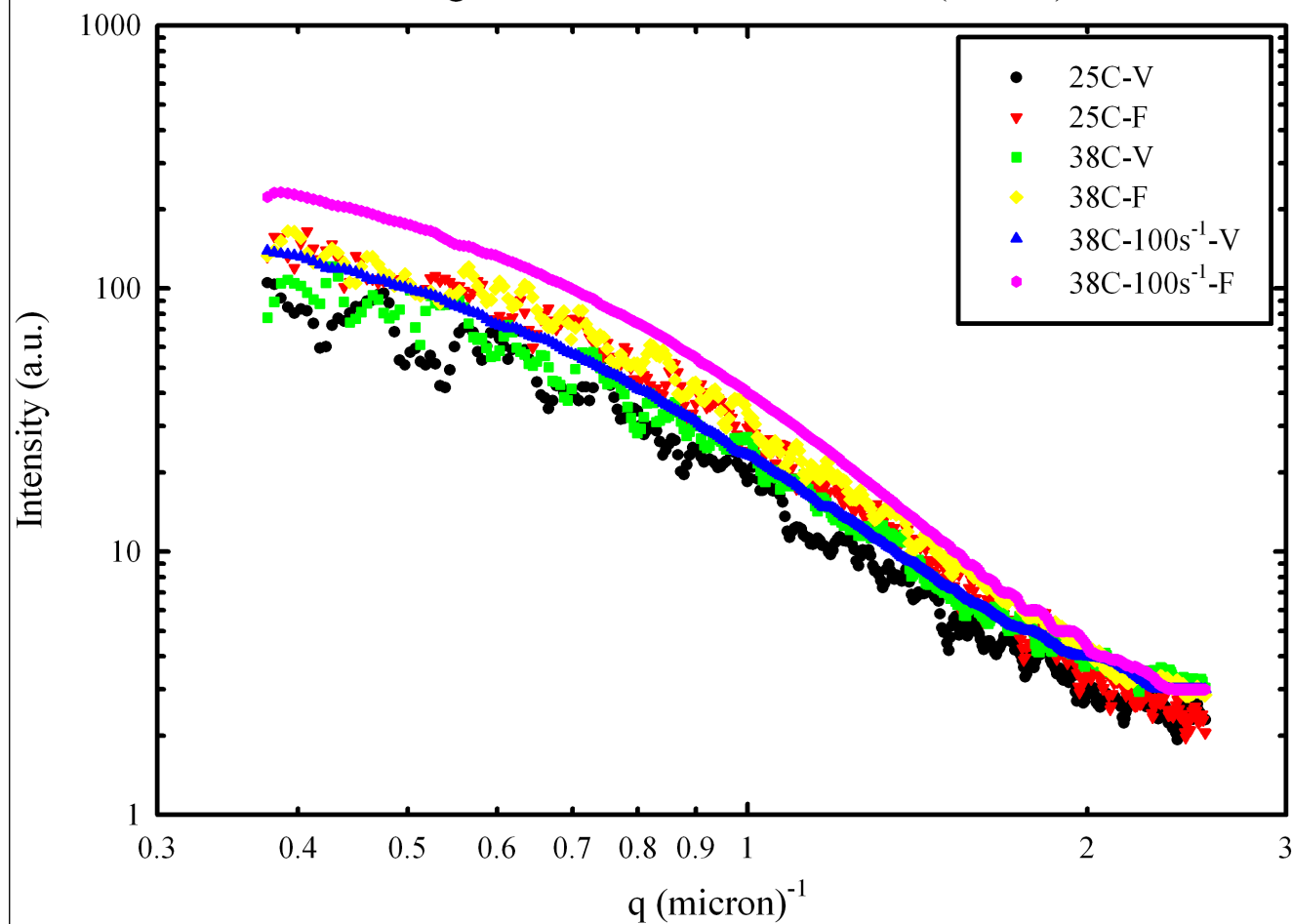
# Scattering



The above graph is for static conditions  
A change from fluid to gel is only seen in SAXS  
and a small change in slope at very low q

# Scattering **shear**

3.5mg/ml fd-NIPAM in 105mM (SALS)



105mM



# Conclusion

1. Gelation temperature and structure : *Can be varied with ionic strength*
2. Power law scaling seen for  $G'$  and *critical strain*. They indicate the difference between the two gels (structure and strength).  $G'$  scaling agrees with theory, but critical strain scaling is underestimated by the theory (by MacKintosh)
3. Gel: Unique  $S$  and  $n$  values have been obtained
  - $S = f(\text{conc. and ionic strength})$  and  $n = f(\text{ionic strength})$
4. Cryo-SEM: *shows structural difference between the gels in two ionic strength?*
5. Scattering technique
  1. Sol to gel transition is seen by SAXS
  2. Only shear densification is seen by SALS (a slope of 2 is seen in both sol and gel states and a slope of 3 under shear)